

# Expert Opinion

1. Introduction
2. Design and development of S-SEDDS formulations
3. Case studies on the development of S-SEDDS formulations
4. Possible drug absorption mechanism from SEDDS, and S-SEDDS formulations and significance of supersaturation-based drug delivery
5. Drug-polymer interaction in sustaining the supersaturated state
6. Expert opinion and conclusions

## Development of supersaturatable self-emulsifying drug delivery system formulations for improving the oral absorption of poorly soluble drugs

Ping Gao<sup>†</sup> & Walter Morozowich

<sup>†</sup> *PGRD, Pfizer, Inc., 301 Henrietta Street, Kalamazoo, MI 49007, USA*

The supersaturatable self-emulsifying drug delivery system (S-SEDDS) represents a new thermodynamically stable formulation approach wherein it is designed to contain a reduced amount of a surfactant and a water-soluble cellulosic polymer (or other polymers) to prevent precipitation of the drug by generating and maintaining a supersaturated state *in vivo*. The S-SEDDS formulations can result in enhanced oral absorption as compared with the related self-emulsifying drug delivery systems (SEDDS) formulation and the reduced surfactant levels may minimise gastrointestinal surfactant side effects.

**Keywords:** bioavailability, lipid-based formulations, microemulsion, oral delivery, poorly soluble drugs, self-emulsifying drug delivery systems, supersaturation, surfactant

*Expert Opin. Drug Deliv.* (2006) 3(1):97-110

### 1. Introduction

#### 1.1 Conventional SEDDS formulations

The development of formulations of highly lipophilic, poorly soluble drugs is often associated with serious challenges with respect to oral absorption. Lipophilic drugs usually have low water solubility and this can result in incomplete oral absorption, low bioavailability and high variability following oral administration. One of the most popular approaches for formulating poorly soluble drugs is the incorporation of the drug into a lipid vehicle containing a surfactant. These formulations represent a unique class of formulations, commonly referred to as self-emulsifying drug delivery systems (SEDDS), and these have been widely explored over the last decade [1-8]. Typically, the SEDDS formulation contains a drug dissolved in a vehicle that is encapsulated in a soft gelatin capsule (softgel), wherein the composition of the vehicle is designed to emulsify rapidly on contact with water and yield a fine oil/water emulsion or microemulsion in the stomach.

SEDDS formulations are commonly developed by an empirical, trial-and-error approach, although some useful guidelines have emerged from the characterisation of successful formulations, such as the cyclosporin A (CsA; Neoral<sup>®</sup>, Sandoz-Novartis) softgel formulation [9-10]. In general, formulation scientists are presented with a series of challenging decisions in the development of SEDDS formulations, beginning with the formulation strategy, excipient selection, solubility and stability assessment, prototype formulation finding and optimisation, scale up and, finally, production of the drug product. The key consideration in the development of a SEDDS formulation of poorly soluble drug is preventing precipitation of the drug following dilution with water. Thus, the drug must remain partitioned within the oil/water emulsion droplets following dilution of the SEDDS formulation with the aqueous medium in the intestine. If the partition coefficient of the drug for the SEDDS emulsion or microemulsion particle is such that the solubility of the drug is

Ashley Publications  
www.ashley-pub.com



exceeded in the aqueous phase, the drug could precipitate following dilution with water, and this could result in poor *in vivo* performance.

### 1.2 Supersaturatable SEDDS formulations

Supersaturatable SEDDS (S-SEDDS) formulations differ from the conventional SEDDS formulations as they contain a reduced amount of surfactant and a polymeric precipitation inhibitor (e.g., water-soluble cellulosic polymers, such as hydroxypropylmethylcellulose [HPMC]), in order to generate and maintain a supersaturated state of the drug following mixing with water.

A high surfactant level is normally employed in conventional SEDDS formulations in order to prevent precipitation of the drug following dilution with water in the gastrointestinal (GI) tract and, in some cases, the surfactants can lead to an increased incidence of GI side effects [11-15].

Increasing the thermodynamic activity of drug formulations and, thereby increasing the bioavailability of poorly soluble drugs, through supersaturation was recognised by Higuchi more than four decades ago [16]. Since then a number of publications have appeared in the literature employing supersaturated formulations as a means of enhancing bioavailability. Most of the work on supersaturation reported in the literature deals with topical delivery [16-27] with less attention on the use of supersaturation for improving the oral delivery of poorly soluble drugs [28-34].

Polyvinylpyrrolidone (PVP) was found to be useful in generating a supersaturated state with a number of poorly soluble drugs [17-21,28,29,35]. Other studies reported the use of the water-soluble cellulosic polymers, such as HPMC [22-25,27,30,31,36], methylcellulose [25], hydroxypropyl methylcellulose phthalate [33,34] and sodium carboxymethylcellulose [37]. The cellulosic polymers are excellent crystal growth inhibitors and are effective in prolonging the supersaturated state of the drugs as shown by a number of *in vitro* studies [21,23,36,37].

Based on the above-cited literature, a promising approach for enhancing the oral bioavailability of poorly soluble drugs is the use of the principle of supersaturation in the development of supersaturatable formulations [38,39]. Supersaturatable formulations differ from supersaturated formulations as they are not thermodynamically stable and, in some cases, the drug can crystallise on storage [40]. In contrast, supersaturatable formulations are thermodynamically stable dosage forms as they yield a supersaturated state only following dilution with water.

## 2. Design and development of S-SEDDS formulations

S-SEDDS formulations of three poorly soluble drugs (paclitaxel, PNU-91325 and Drug X) were developed and their *in vitro* behaviour was characterised, and the oral bioavailability was determined. The *in vitro* test methodology employing biorelevant conditions is useful in guiding the development of

the S-SEDDS formulations, as well as in the development of *in vitro/in vivo* relationships.

Prompted by the biorelevant dissolution system reported by Tang *et al.* [41] and the importance of GI physiology and dosage form performance, a small-scale *in vitro* release/precipitation test was developed for evaluating and optimising the S-SEDDS formulations of the three poorly soluble drugs. The biorelevant *in vitro* release/precipitation test method consisted of simulated gastric fluid (SGF) containing 0.01 M HCl and 0.15 M NaCl (pH 2) stirred at 50 rpm at 37°C [38,39]. The total volume of the medium is 50 – 100 ml, which is the approximate volume of the stomach fluid when swallowing a tablet or capsule dosage form with 2 – 3 oz of water.

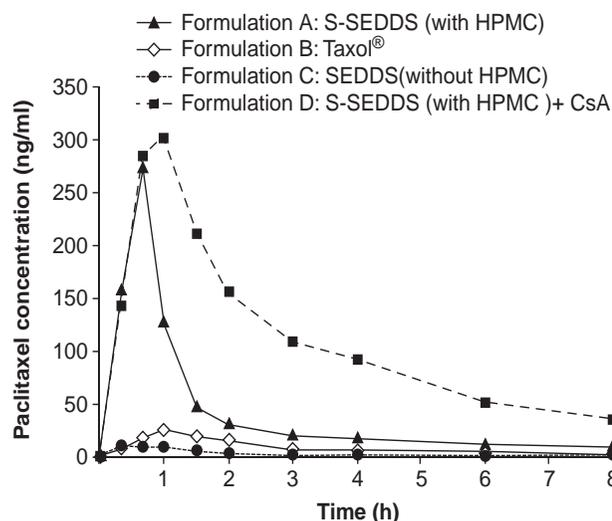
The *in vitro* release/precipitation test is conducted by placing a unit dose of the prototype S-SEDDS formulation or related formulation in the stirred test medium at 37°C and withdrawing samples from the test medium as a function of time followed by filtration (0.8 µm), and the determination of the total drug concentration in the filtrate using high-performance liquid chromatography. The apparent drug concentration following filtration of the S-SEDDS formulation during the test provides the total concentration of the drug in the filtrate, which consists of free drug along with some emulsion, micelle or solid drug particles with a particle size < 0.8 µm generated within in the release/precipitation test medium [38,39]. The *in vitro* release/precipitation test and the resulting apparent drug concentration versus time plots were employed in guiding the development of the S-SEDDS formulations, as illustrated by the three case studies described below.

## 3. Case studies on the development of S-SEDDS formulations

### 3.1 Paclitaxel

Paclitaxel is an antitumour agent that is widely used in the treatment of advanced breast and ovarian cancer. Paclitaxel has a molecular weight of 853 Da and a low solubility in water (< 1 µg/ml), as well as a low solubility in common pharmaceutical vehicles [42,43]. The currently marketed intravenous formulation of paclitaxel (Taxol®, Bristol-Myers Squibb) contains paclitaxel 6 mg/ml, Cremophor EL 527 mg/ml (polyoxyethylenated castor oil) and 49.7% (v/v) of dehydrated ethanol [44]. Intravenous administration of paclitaxel using this formulation is associated with side effects that are attributed to the surfactant Cremophor EL and these side effects can be controlled by the coadministration of antihistamine [43,45-47]. The oral bioavailability of paclitaxel using the Taxol IV formulation is extremely low (< 2%) when administered orally to animals and humans [48-50]. Oral coadministration of the Taxol IV formulation along with CsA, an inhibitor of P-glycoprotein and cytochrome P450 3A enzymes, resulted in a sevenfold increase in the plasma area under the curve (AUC) value for paclitaxel in humans [48-51].

The mean plasma concentration of paclitaxel obtained in rats with the SEDDS and S-SEDDS formulations and



**Figure 1. Mean plasma concentration-time profiles of paclitaxel in rats after oral administration using four formulations.** Reprinted with permission from GAO P, RUSH RD, PFUND WP *et al.*: Development of a S-SEDDS formulation of paclitaxel with improved oral bioavailability. *J. Pharm. Sci.* (2003) **92**(12):2395-2407.

CsA: Cyclosporin A; HPMC: Hydroxypropylmethylcellulose; SEDDS: Self-emulsifying drug delivery systems; S-SEDDS: Supersaturatable self-emulsifying drug delivery system.

Taxol are plotted in **Figure 1** [39]. The pharmacokinetic parameters corresponding to these treatment groups are summarised in [39]. The rank order of the mean total exposure as given by the  $AUC_{0-\infty}$  for the four formulations is S-SEDDS + CsA > S-SEDDS >> Taxol ~ SEDDS.

The difference in the pharmacokinetic profiles exhibited by the SEDDS and S-SEDDS formulations is intriguing because these two formulations differ only in the content of HPMC; 0 and 5%, respectively. Thus, the SEDDS formulation (without HPMC) showed a very low maximum concentration ( $C_{max}$ ) of only 13.1 ng/ml and an oral bioavailability of 0.9%, whereas the S-SEDDS formulation (with HPMC) resulted in an impressive 20-fold increase in  $C_{max}$  (~ 277 ng/ml) and a 10-fold increase in the oral bioavailability; namely 9.5% (**Figure 1**). The S-SEDDS formulation with the addition of CsA and HPMC showed similar absorption kinetics; however, the slower elimination kinetics, resulted in a twofold increase in the oral bioavailability over that of the S-SEDDS formulation containing only HPMC. As discussed in [39], the rat bioavailability results indicate that the higher paclitaxel solution concentration generated by the S-SEDDS formulation in the *in vitro* release/precipitation test as a result of supersaturation is responsible for the enhanced oral bioavailability of paclitaxel.

The Taxol formulation generates a nearly transparent solution (particle size < 30 nm) following dilution with water, and no precipitation of paclitaxel is observed for several days after dilution of the formulation with water. The inhibition of precipitation of the drug on dilution of this formulation

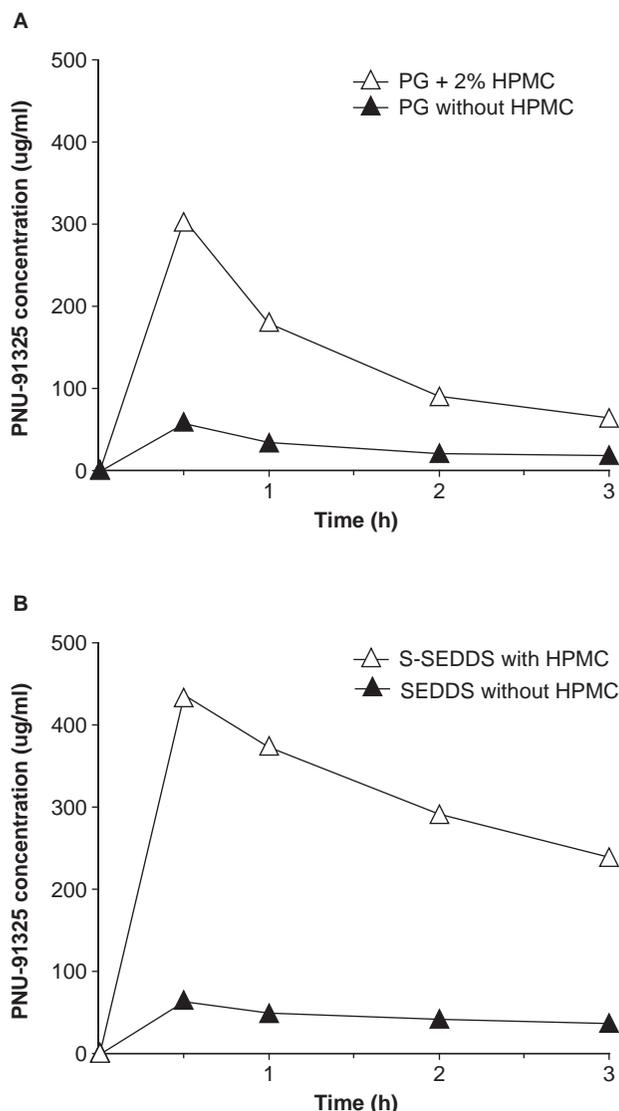
with water is due to the high Cremophor EL content in the formulation (weight ratio of Cremophor EL:paclitaxel was 88:1) and this results in complete solubilisation of paclitaxel by the Cremophor micelles. Apparently, the free-drug concentration does not exceed the solubility of paclitaxel in water [39]. It is noteworthy that the Taxol formulation (**Figure 1**) produced a low mean  $C_{max}$  (~ 30 ng/ml) and a low oral bioavailability (~ 2%). Thus, the poor oral exposure of paclitaxel from the Taxol formulation in rats is probably due to the excess Cremophor, resulting in sequestration of the paclitaxel within the micelle and reducing the free-drug concentration in the formulation [39], which is consistent with similar reports in the literature [52,53].

### 3.2 PNU-91325

PNU-91325 is a lipophilic drug with a calculated Log P partition coefficient (CLogP) of 2.8 [38]. This molecule shows a U-shaped pH-solubility dependency, indicating two  $-\log_{10}$  dissociation constant for an acid ( $pK_a$ ) values (a basic  $pK_a$  of 2.61 and an acidic  $pK_a$  of 6.85). PNU-91325 shows an almost constant solubility of ~ 6  $\mu\text{g/ml}$  within the physiological pH range of 2 – 7.

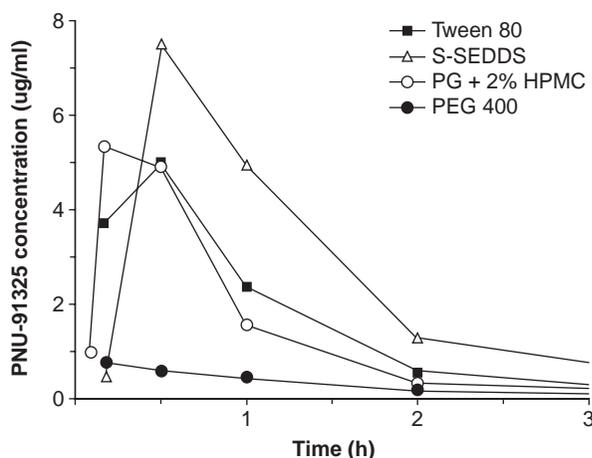
The difference in the *in vitro* release/precipitation profile between the two formulations, with and without HPMC, clearly indicates that a small amount of HPMC plays a critical role in achieving a supersaturated state with PNU-91325 by retarding drug precipitation and, in addition, sustaining the high supersaturated drug concentration. As shown in **Figure 2A**, in the absence of HPMC the PNU-91325 concentrations were low following administration of a propylene glycol (PG) formulation, whereas a much higher concentration was observed when 2% HPMC was incorporated in the same PG formulation. Similarly, the PNU-91325 concentrations from an S-SEDDS formulation with HPMC increased by approximately fivefold as compared with those of the same SEDDS formulation without HPMC during the time course of 0 – 3 h.

The mean oral plasma concentrations of PNU-91325 in beagle dogs (n = 4, crossover) are plotted in **Figure 3** as obtained with the Tween 80 formulation, the polyethylene glycol (PEG) 400 formulation, the S-SEDDS formulation and the PG plus 2% HPMC (S-cosolvent) formulation [38]. The mean dose, mean  $AUC_{0-\infty}$ ,  $C_{max}$ , and the estimated absolute bioavailability values observed with each of the formulations are reported in [38]. The low oral bioavailability observed with the PEG 400 solution of PNU-91325 is probably due to the rapid precipitation of the drug *in vivo*. As indicated by the above *in vitro* release/precipitation test data, the PG plus 2% HPMC (S-cosolvent) formulation of PNU-91325 yielded a significantly higher apparent drug concentration (**Figure 2B**). This formulation resulted in an approximately sevenfold higher  $C_{max}$  (6.04  $\mu\text{g/ml}$ ) and a fivefold higher oral bioavailability (~ 60%) as compared with the PEG 400 formulation that showed a lower  $C_{max}$  (0.88  $\mu\text{g/ml}$ ) and a lower oral bioavailability (~ 12%).



**Figure 2. Apparent concentration-time profiles of PNU-91325 observed in the *in vitro* dissolution/precipitation test using formulations with or without HPMC.** Copyright (2004) from GAO P, GUYTON ME, HUANG T, BAUER JM, STEFANSKI KJ, LU Q: Enhanced oral bioavailability of a poorly water soluble drug PNU-91325 by supersaturatable formulations. *Drug Devel. Ind. Pharm.* (2004) **30**(2):221-229. Reprinted by permission of Taylor & Francis, Inc., <http://www.taylorandfrancis.com>.  
HPMC: Hydroxypropylmethylcellulose; PG: Propylene glycol; SEDDS: Self-emulsifying drug delivery systems; S-SEDDS: Supersaturatable self-emulsifying drug delivery system.

The PNU-91325 S-SEDDS formulation showed an oral bioavailability of ~ 76%, which is higher than the bioavailability of ~ 68% observed with the Tween 80 formulation. As the formulation compositions were reported in Table 1 in [38], the weight ratio of drug:Cremophor EL is 1:7.5 in the S-SEDDS formulation, whereas the weight ratio of



**Figure 3. Mean plasma concentration-time profiles of PNU-91325 in dogs (n = 4, crossover) using four PNU-91325 formulations.** Copyright (2004) from GAO P, GUYTON ME, HUANG T, BAUER JM, STEFANSKI KJ, LU Q: Enhanced oral bioavailability of a poorly water soluble drug PNU-91325 by supersaturatable formulations. *Drug Devel. Ind. Pharm.* (2004) **30**(2):221-229. Reprinted by permission of Taylor & Francis, Inc., <http://www.taylorandfrancis.com>.  
HPMC: Hydroxypropylmethylcellulose; PEG: polyethylene glycol; PG: Propylene glycol; S-SEDDS: Supersaturatable self-emulsifying drug delivery system.

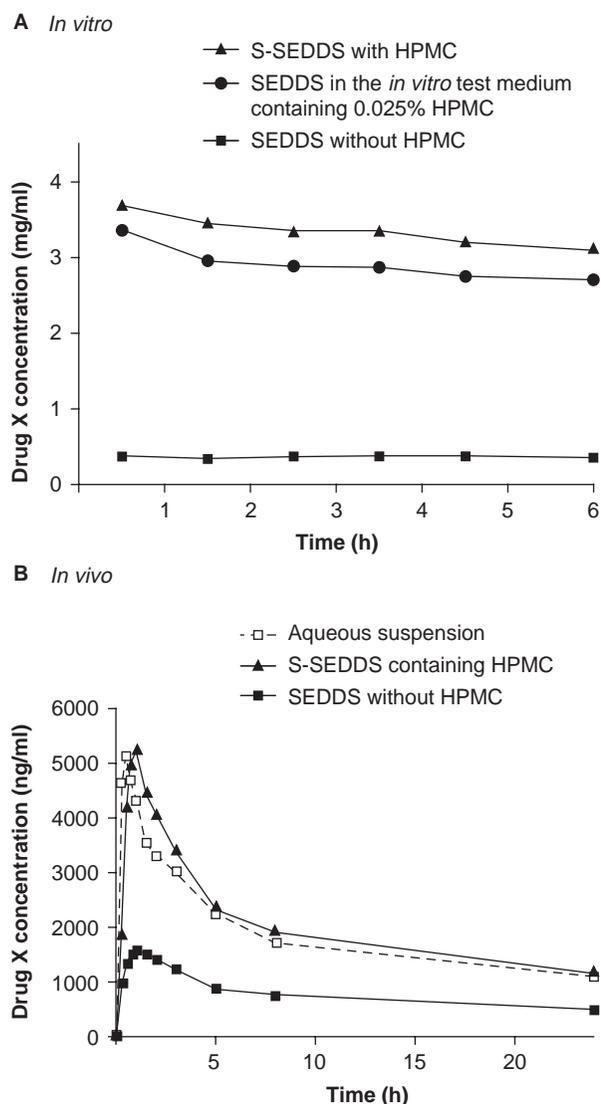
drug:Tween 80 is 1:39 in the Tween 80 formulation. Clearly, the surfactant level is about fivefold higher in the Tween 80 formulation as compared with the S-SEDDS formulation. The S-SEDDS formulation yielded a supersaturated state following dilution as evidenced by the decline in the apparent drug concentration with respect to time (Figure 2B) and by the observation of crystal formation in the *in vitro* release/precipitation test medium [38].

### 3.3 Drug X

Formulations of Drug X were under development for preclinical and clinical evaluation. Drug X has a log P of ~ 3.5, a water solubility of only ~ 5 µg/ml in the physiological pH range of 2 – 7 and it is nonionisable in this pH range. A human oral pharmacokinetic study using Drug X showed slow and incomplete oral absorption using the drug powder in a gelatin capsule, whereas rapid and more complete absorption was found with a small particle size aqueous suspension of Drug X.

#### 3.3.1 S-SEDDS formulations containing suspended HPMC powder

The *in vitro* release/precipitation test used 1 g of the S-SEDDS formulations containing 200 mg of Drug X filled into two hard gelatin capsules (0.5 g/capsule) and 50 ml of SGF to yield a theoretical concentration of Drug X in the test medium of 4 mg/ml. The apparent Drug X concentration found with the SEDDS formulation (without HPMC) in the



**Figure 4. A. Apparent concentration–time profiles of Drug X observed in the *in vitro* release/precipitation test using the same SEDDS formulation with and without HPMC.** All formulations were filled into hard gelatin capsules. **B. Mean plasma concentration profiles of Drug X in the dogs *in vivo* ( $n = 6$ , crossover) using the SEDDS and S-SEDSS formulations as compared with an aqueous suspension formulation [54].**

HPMC: Hydroxypropylmethylcellulose;  
 SEDDS: Self-emulsifying drug delivery systems;  
 S-SEDSS: Supersaturatable self-emulsifying drug delivery system.

*in vitro* release/precipitation test is plotted in Figure 4A. The concentration of Drug X in the medium was  $\sim 0.3$  mg/ml at the first time point (0.5 h) and this remained unchanged over the entire 6-h test period. A white precipitate of Drug X was observed with the SEDDS formulation in the release/precipitation test medium at 0.5 h and the precipitate was found to be crystalline as shown by polarised light microscopy. The formation of drug crystals in the test medium indicates that

the solution was supersaturated with respect to the crystalline form of Drug X. In contrast, a markedly higher concentration of Drug X ( $\sim 2.7 - 3.5$  mg/ml) was observed with the same SEDDS formulation in SGF containing 0.025% (Figure 4A). The presence of HPMC at the remarkably low concentration of only 0.25 mg/ml (0.025%) in the test medium is sufficient to generate and maintain the supersaturated state with Drug X for at least 6 h under the conditions of the release/precipitation test.

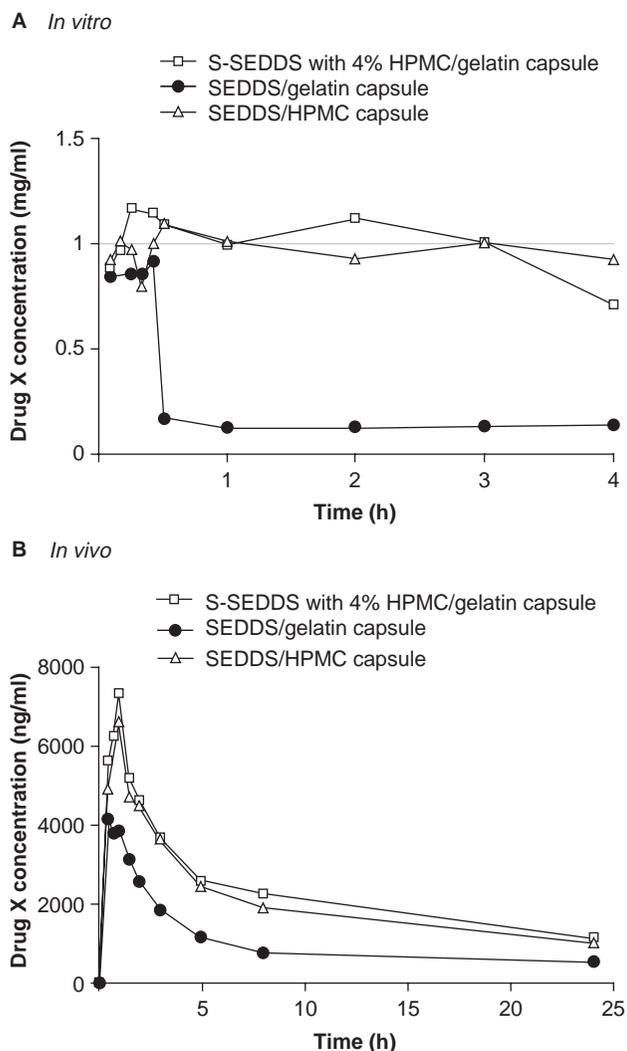
Little precipitation of Drug X was observed over the 6-h test period in which the S-SEDSS formulation and the Drug X concentration was  $\sim 3 - 3.5$  mg/ml. This is similar to the above case where the HPMC was dissolved in the SGF medium prior to the release/precipitation study. The apparent Drug X concentration from the S-SEDSS formulation in the release/precipitation test was  $\sim 10$ -fold higher than the SEDDS formulation without HPMC in the release medium.

The *in vivo* pharmacokinetics of both the SEDDS and the S-SEDSS formulations of Drug X were evaluated after oral administration in dogs in comparison with an aqueous suspension of Drug X. Figure 4B shows that the mean plasma concentration profile of Drug X obtained after dosing the S-SEDSS formulation (with 4.4% HPMC, w/w) is about threefold higher in the  $C_{max}$  value, and the AUC is 2.5-fold larger as compared with that of the same SEDDS formulation without HPMC. This clearly indicates that the S-SEDSS formulation containing HPMC results in an increase in both the  $C_{max}$  and the extent of absorption of Drug X. The aqueous suspension and the S-SEDSS formulation showed a similar pharmacokinetics profile in dogs with slightly higher  $C_{max}$  and AUC values for the S-SEDSS formulation.

### 3.3.2 S-SEDSS formulations in a HPMC capsule

The use of a HPMC capsule shell was explored as an alternate approach for incorporating HPMC into the S-SEDSS formulation. The same SEDDS liquid fill that was used in the above studies was filled into HPMC capsules (Quali-V capsules, Shionogi; capsule shell weight  $\sim 90$  mg). Three dosage forms were selected for comparison in the *in vitro* release/precipitation test; a SEDDS liquid formula filled into hard gelatin capsules, the same SEDDS liquid formula with 44 mg of HPMC powder (suspension) in a hard gelatin capsule and the same SEDDS liquid formula filled into a HPMC capsule.

The SEDDS liquid formula in all three formulations was identical. Figure 5A shows the apparent drug concentrations of Drug X as a function of time obtained with these three dosage forms in the *in vitro* release/precipitation test. As expected, the SEDDS liquid in the hard gelatin capsule showed a Drug X concentration of  $\sim 1$  mg/ml initially (at 15 min) in the release test. However, the Drug X solution concentration rapidly decreased to  $\sim 0.2$  mg/ml within 30 min and the concentration remained unchanged. In contrast, the 1-g SEDDS formulation containing HPMC 44 mg suspended in a hard gelatin capsule showed an almost constant drug concentration of



**Figure 5. A. Apparent concentration–time profiles of Drug X observed in the *in vitro* release/precipitation test using the three formulations with different capsule shells as indicated. B. Mean plasma concentration profiles of Drug X in the dogs from the three formulations ( $n = 6$ , crossover) [54].**

HPMC: Hydroxypropylmethylcellulose;  
 SEDDS: Self-emulsifying drug delivery systems;  
 S-SEDDS: Supersaturatable self-emulsifying drug delivery system.

~ 1 mg/ml over the entire 4-h period (Figure 5A). The SEDDS liquid filled into a HPMC capsule showed essentially the same concentration–time profile as the SEDDS formulation containing suspended HPMC powder filled into gelatin capsules. In both cases, where HPMC was present either as suspended powder in the SEDDS liquid or as HPMC provided by the capsule shell, the Drug X concentration was maintained at a level about fivefold higher than that of the SEDDS liquid alone in a hard gelatin capsule. This data clearly indicates that a HPMC capsule acts in a similar way to HPMC suspended within the SEDDS liquid with respect to achieving and maintaining the supersaturated state with Drug X and, thus, both of these are S-SEDDS formulations.

An oral bioavailability study was conducted in dogs ( $n = 6$ , crossover) with the three SEDDS formulations. The mean plasma concentration–time profiles of Drug X are plotted in Figure 5B. As expected, the SEDDS formulation of Drug X in the gelatin capsule showed a low  $C_{max}$  and a low AUC. However, the plasma concentration–time profiles observed with the two S-SEDDS formulations, consisting of the SEDDS formulation containing HPMC, and the SEDDS formulation filled into HPMC capsules, were almost superimposable and the resulting  $C_{max}$  and AUC values were approximately twofold higher than that of the SEDDS liquid without HPMC in the gelatin capsule. The *in vivo* behaviour of the three formulations is in accord with the *in vitro* release/precipitation test results.

In summary, Drug X in S-SEDDS formulations containing HPMC either as suspended powder or as HPMC in the capsule shell results in achieving and maintaining a supersaturated Drug X solution following contact with water, and this results in higher *in vivo* oral bioavailability in dogs. Further evaluation in the clinic is discussed in the following section.

### 3.3.3 Clinical evaluation of the S-SEDDS formulation

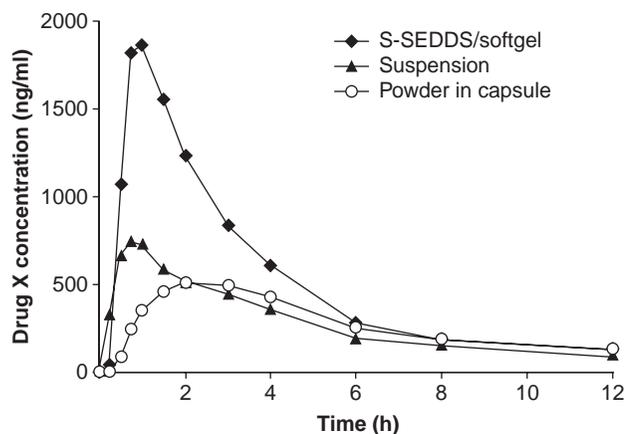
S-SEDDS softgel formulations of Drug X were evaluated in fasted humans in comparison with two other formulations, namely a bulk drug powder formulated in a gelatin capsule and an aqueous suspension of Drug X (23 subjects, crossover)

The clinical study results obtained with the three dosage forms of Drug X are shown in Figure 6 where the mean  $C_{max}$ , maximum time ( $T_{max}$ ) and AUC values are taken from [54]. The bulk drug powder formulation in the gelatin capsule showed the lowest  $C_{max}$  (621 ng/ml) and the aqueous suspension of the drug showed a slightly higher  $C_{max}$  (804 ng/ml). In contrast, the S-SEDDS softgel containing suspended HPMC showed the highest  $C_{max}$  (2061 ng/ml), which is an impressive 331% increase in  $C_{max}$  and a 40% increase in the AUC as compared with the Drug X powder formulation in the gelatin capsule. The highest  $C_{max}$  and the largest AUC along with the shortest  $T_{max}$  (~ 1 h) were observed with the S-SEDDS softgel containing HPMC indicating more rapid and more complete absorption.

## 4. Possible drug absorption mechanism from SEDDS and S-SEDDS formulations, and significance of supersaturation-based drug delivery

### 4.1 Drug absorption from the SEDDS and S-SEDDS formulations

Drugs with water solubilities < ~ 1 – 50  $\mu\text{g/ml}$  frequently show incomplete oral absorption when the dose is  $\geq 50$  mg [55–59]. However, there are a number of other extremely insoluble and highly lipophilic compounds that have solubilities that are orders of magnitude < 1 – 50  $\mu\text{g/ml}$  and yet, they are absorbed orally [60–62]. The early work of Børgstrom *et al.* [63,64] and later Carey *et al.* [65–67], as well as many others [68–99], showed that it is the bile acid mixed micelle (BAMM)



**Figure 6.** Human bioavailability study (subject number = 23, cross-over) with three formulations of Drug X: formulated Drug X powder/hard gelatin capsule, an aqueous suspension, and a S-SEDDS formulation/softgel [54].

S-SEDDS: Supersaturatable self-emulsifying drug delivery system.

in the fed state and the bile acid (BA) micelle in the fasted state that constitute the endogenous surfactant system that is responsible for the presentation of extremely lipophilic compounds and drugs to the surface of the enterocyte brush border region where transfer of the compound from the BA/BAMM particle to the glycocalyx can occur by collisional contact [100-103].

Cholesterol with a ClogP of 12 and a water solubility of  $\sim 10$  ng/ml is efficiently absorbed from the intestine by initial presentation of cholesterol in the BAMM particles to the enterocyte brush border mucosa with subsequent collisional transfer to the glycocalyx [63-68]. Many other extremely insoluble and lipophilic compounds are absorbed more efficiently in the fed state due to the high concentration of BAMM in the intestine in this state [55]. The BAMM system in the fasted state is more effective in solubilising drugs than the BA system in the fasted state because of the higher total micellar concentration of BAMM as compared with that of BA in the fasted state.

Lipophilic drugs are known to partition into BA/BAMM systems [99,104-106] and predictive relationships have been reported [106]. Highly lipophilic compounds can equilibrate between populations of liposomes by collisional contact, whereas less lipophilic compounds can equilibrate between populations of liposomes via the compound that is dissolved in the aqueous medium [107].

Based on the above, it appears that drugs can be delivered to the intestinal enterocyte brush border region by the aqueous diffusion pathway, as well as by the BA/BAMM pathway. The delivery of cholesterol from the BAMM to the enterocyte surface occurs via collisional transfer [88-90,93,94,96-98,108-113,201].

**Figure 7** shows a possible scheme for the presentation of poorly soluble lipophilic drugs from SEDDS and S-SEDDS formulations to the intestinal enterocyte brush border by three possible pathways; the aqueous pathway; by

mimicking the BA/BAMM pathway; or by equilibrating with the BA/BAMM pathway.

**Figure 7** shows that the emulsion or microemulsion generated in the intestinal lumen from a SEDDS/S-SEDDS soft-gel formulation can undergo lipolysis or transfer of the excipients to the enterocyte (path A). The drug in the remnant emulsion/microemulsion or the resulting micelle that is generated from the SEDDS/S-SEDDS formulation can undergo collisional transfer to the glycocalyx (path B). Alternatively, the drug in the aqueous medium can become absorbed directly by the well-known passive diffusion aqueous pathway (path C). The free drug in the aqueous medium could equilibrate with the BAMM or remnant BAMM with collisional transfer of the drug to the glycocalyx (path D) without requiring water solubility.

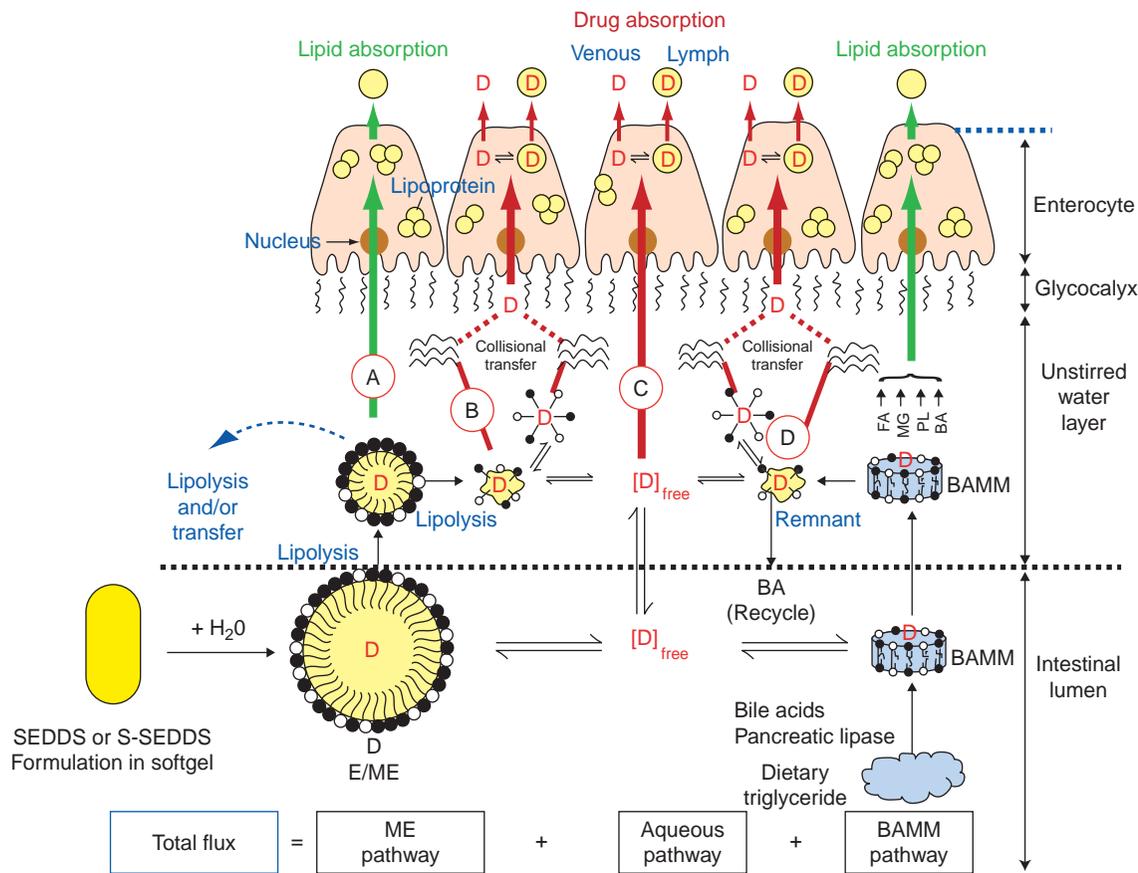
After absorption into the enterocyte, the drug could diffuse across the enterocyte or, if the drug is highly lipophilic, the drug could partition into the chylomicrons in the fed state with subsequent transfer to the lymphatics and ultimately to the systemic venous circulation [5,8,114,115]. Moderately lipophilic drugs could escape from the chylomicrons by partitioning or collisional transfer with subsequent entry into the venous system.

Lipid digestion can be important in the absorption of highly lipophilic drugs because of their probable high partition affinity for the emulsion/microemulsion particle. Lipids, such as long-chain fatty acid triglycerides, must undergo lipolysis before absorption of the resulting fatty acids can occur. The lipid digestion process and the subsequent absorption of some lipophilic drugs can be adversely affected by the presence of surfactants in the formulation [58], and by a large droplet size for the emulsion/microemulsion particle formed following exposure to GI fluids.

#### 4.2 Significance of supersaturation in enhancing drug absorption

Supersaturation was shown to occur in the authors' *in vitro* release/precipitation test, wherein the drug filtrate concentration from a S-SEDDS formulation of PNU-91325 showed a maximum value of  $\sim 420$   $\mu\text{g/ml}$  at 30 min (**Figure 2B**, S-SEDDS with HPMC). However, the SEDDS formulation of PNU-91325, without HPMC, yielded a drug filtrate concentration of only  $\sim 30$   $\mu\text{g/ml}$  at 3 h in the release/precipitation test and, thus, the approximate degree of apparent supersaturation with the S-SEDDS formulation of PNU-91325 is 420/30 or about 14. The  $\sim 14$ -fold degree of apparent supersaturation observed with the S-SEDDS formulation of PNU-91325 suggests that intestinal absorption should be markedly increased as a result of the supersaturated state. Initially, the test medium did not show evidence of drug crystals under the microscope; however, on long standing, crystals were readily seen, thus indicating that the medium was supersaturated.

The failure to provide high oral exposure of paclitaxel with the Taxol formulation is significant in that the common



**Figure 7. Proposed intestinal drug absorption via the alternate pathways for presentation of drugs to the intestinal glycocalyx.** Left to right; **A.** Lipids in E/ME can be absorbed upon lipolysis; **B.** the remnant E/ME particle, with probable enhanced thermodynamic potential, can transfer the drug to the glycocalyx via collisional transfer; **C.** the free drug in equilibrium with the E/ME and the remnant E/ME can be absorbed by passive diffusion; **D.** the free drug could equilibrate with the BAMM and then undergo collisional transfer with the glycocalyx.

BA: Bile acid; BAMM: Bile acid mixed micelle; D: Drug; E: Emulsion; FA: Fatty acid; MG: Monoglyceride; PL: Phospholipid; SEDDS: Self-emulsifying drug delivery systems; S-SEDDS: Supersaturatable self-emulsifying drug delivery system; ME: Microemulsion.

practice of formulating poorly soluble drugs with high concentrations of surfactants inevitably results in the reduction in the free-drug concentration or the thermodynamic activity. It is well known that micelle solubilisation of poorly soluble, lipophilic drugs can result in a low free-drug concentration, as given by the partition coefficient, and the low free-drug level can result in a reduced rate and extent of absorption [13-15]. The work by Poelma *et al.* [13,14] is noteworthy in that the kinetics of the intestinal absorption of griseofulvin in rats was found to be directly related to the concentration of the free griseofulvin level found in the aqueous solutions containing Tween 80 (a surfactant) [13]. These workers also showed a reduction in the rate of absorption of griseofulvin from the small intestine of rats in the presence of 10 – 20 mM taurocholate, and reduction was directly related to the reduced free drug level [14]. Likewise, Chiu *et al.* [15] showed that the presence of surfactants (Cremophor EL, RH40 and VE-TPGS with surfactant concentrations at 0.02% weight/volume or higher) significantly decreased the apparent permeability of CsA in Caco-2 cells, and the

magnitude of the decrease in permeability (2- to 12-fold) was directly related to the surfactant concentration. Reduction in the apparent permeability of CsA in the presence of surfactants decreased the free drug levels of CsA in the solution as a result of micellar solubilisation.

In conclusion, the mechanism responsible for the enhanced intestinal absorption of poorly soluble drugs from S-SEDDS formulations containing HPMC is probably due to enhanced presentation of the drug to the enterocyte brush border region by the aqueous pathway due to the increased free-drug levels achieved by supersaturation, in addition to presentation of the drug to the enterocyte brush border by mimicking, or equilibrating with, the BA/BAMM pathway.

### 5. Drug-polymer interaction in sustaining the supersaturated state

Although there have been many reports in the literature on inhibition of crystallisation by HPMC and other polymeric

materials [16-37] with pharmaceutical substances, the underlying mechanism of inhibition is complex and is rarely illustrated or studied.

The ability to generate a supersaturated state with HPMC with the S-SEDDS formulations may be due to the formation of a widely spaced cellulosic-polymer network that is created by the HPMC chains in water. According to the literature, solutions of HPMC consist of 'cellulosic bundles resulting in a tenuous network of swollen clusters with hydrophobic substituents surrounded by sheaths of structured water' [116,117]. Studies on the mechanism responsible for inhibiting crystallisation of drugs in aqueous solutions containing HPMC suggests that the HPMC polymer chain may inhibit nucleation, as well as crystal growth by adsorption of the HPMC molecules onto the surface of the nuclei, or onto the surface of crystals [20,28,118]. The general applicability of cellulosic polymers in inhibiting crystallisation of many pharmaceutical substances is widely reported [21-23,25,27,31,33,34,36].

Simonelli *et al.* [28] suggested that the polymer at the crystal surface forms a net-like structure, which allows the drug to grow out in finger-like protrusions leading to a growth with a rough surface. Ziller *et al.* [118] suggests that the polymer inhibits the introduction of drug molecules from solution into the crystal lattice by occupying adsorption sites and, thus, the adsorbed polymer forms a mechanical barrier that inhibits crystallisation. Raghavan *et al.* [20] proposed that the mechanism of nucleation and growth is based on the interaction between the drug and the polymer molecules through hydrogen bonding. In their work, they pointed out that the adsorption of the polymer substance on the nuclei surface of hydrocortisone acetate occurs through hydrogen bonding. The hydrodynamic boundary layer surrounding the crystal, resulting from adsorption of the polymer molecules onto the crystal surface, leads to crystal growth inhibition as well as habit modification of the crystals.

The general applicability of HPMC polymer in inhibiting crystallisation of many pharmaceutical substances is

widely reported [22-25,27,30,31,36], suggesting that a nonspecific mechanism may be involved in the inhibition of crystallisation. Understanding the mechanism for inhibition of crystallisation could aid the selection of other effective precipitation inhibitors and this could enhance the application of supersaturatable formulations

## 6. Expert opinion and conclusions

The S-SEDDS formulation is designed to contain both a reduced amount of surfactant(s) and a polymeric precipitation inhibitor (e.g., water-soluble cellulosic polymers, such as HPMC), and generate and maintain a supersaturated state of the drug following mixing with water. The three case studies with paclitaxel, PNU-91325 and the exploratory drug Drug X, clearly show that the new S-SEDDS formulation approach provides higher oral bioavailability as compared with that of the conventional SEDDS formulations. Increasing the free-drug concentration by generating and maintaining a supersaturated state in the intestine is a means of overcoming the solubility limited absorption problems typical of poorly soluble drugs.

In addition to the potential for enhanced oral bioavailability, the S-SEDDS formulations could provide rapid onset such as in acute pain relief, migraine and other therapies. Rapid onset is difficult to achieve with poorly soluble drugs because of their low solubility and slow dissolution of the crystalline solids. Another advantage of S-SEDDS formulations is their potential for reducing the extent of GI side effects typically seen with SEDDS formulations due to the reduced surfactant levels in the S-SEDDS formulations.

Further exploration of the S-SEDDS formulation technology including an understanding of the precipitation kinetics and the mechanism for inhibiting drug precipitation with polymeric substances could lead to improved supersaturatable formulation strategies with enhanced oral absorption of poorly soluble drugs.

## Bibliography

Papers of special note have been highlighted as either of interest (•) or of considerable interest (••) to readers.

- GURSOY RN, BENITA S: Self-emulsifying drug delivery systems (SEDDS) for improved oral delivery of lipophilic drug. *Biomed. Pharmacother.* (2004) **58**(3):173-82.
- WASAN KM: Formulations and physiological and biopharmaceutical issues in the development of oral lipid-based drug delivery systems. *Drug Devel. Ind. Pharm.* (2001) **27**(4):267-276.
- BAGWE RP, KANICKY JR, PALLA BJ, PATANJALI PK, SHAH DO: Improved drug delivery using microemulsions: rationale, recent progress, and new horizons. *Crit. Rev. Ther.* (2001) **18**(1):77-140.
- HAUSE D: Lipid based systems for oral drug delivery: enhancing the bioavailability of poorly soluble drugs. *Am. Pharm. Rev.* (2002) Nov/Dec:22-26.
- CHARMAN CH: Lipids, lipophilic drugs and oral drug delivery - some emerging concepts. *J. Pharm. Sci.* (2000) **89**:967-978.
- POUTON CW: Lipid formulations for oral administration of drugs: non-emulsifying, self-emulsifying and 'self-microemulsifying' drug delivery systems. *Eur. J. Pharm. Sci.* (2000) **11**(Suppl. 2):S93-S98.
- HUMBERSTONE AJ, CHARMAN WN: Lipid-based vehicles for oral delivery of poorly soluble drugs. *Adv. Drug Deliv. Rev.* (1997) **25**:103-128.
- POUTON CW: Formulations of self-emulsifying drug delivery systems. *Adv. Drug Deliv. Rev.* (1997) **25**:47-48.
- VONDERSCHER J, MEINZER A: Rationale for the development of Sandimmune Neoral. *Transplant. Proc.* (1994) **26**(5):2925-2927.
- The scientific rationale for developing the microemulsion formulation of cyclosporin was revealed.

10. MEINZER A, MUELLER E, VONDERSCHER J: Microemulsion-A suitable galenical approach for the absorption enhancement of low soluble compounds? *B. T. Gattafosse* (1995) **N88**:21-27.
11. WIGNOT TM, STEWART RP, SCHRAY KJ, DAS S, SIPOS T: *In vitro* studies of the effects of HAART drugs and excipients on activity of digestive enzymes. *Pharm. Res.* (2004) **21**(3):420-427.
12. SHERMAN DS, FISH DN: Management of protease inhibitor associated diarrhea. *Clin. Infect. Dis.* (2000) **30**:908-914.
13. POELMA FG, BREAS R, TUKKER JJ, CROMMELIN DJ: Intestinal absorption of drugs. The influence of mixed micelles on the disappearance kinetics of drugs from the small intestine of the rat. *J. Pharm. Pharmacol.* (1991) **43**(5):317-324.
14. POELMA FG, BREAS R, TUKKER JJ, JOSEF J: Intestinal absorption of drugs. III. The influence of taurocholate on the disappearance kinetics of hydrophilic and lipophilic drugs from the small intestine of the rat. *Pharm. Res.* (1990) **7**(4):392-397.
- **The reduction in the absorption kinetics of griseofulvin in the presence of taurocholate correlated well with the reduction of the drug-free fraction in solution due to micellar solubilisation.**
15. CHIU YY, HIGAKI K, NEUDECK BL, BARNETT JL, WELAGE LS, AMIDON GL: Human jejunal permeability of cyclosporin A: influence of surfactants on P-glycoprotein efflux in Caco-2 cells. *Pharm. Res.* (2003) **20**(5):749-756.
16. HIGUCHI T: Physical chemical analysis of the percutaneous absorption process. *J. Soc. Cosmet. Chemists* (1960) **11**:85-97.
- **First theoretical basis for enhancing skin absorption of drugs by supersaturated solutions.**
17. MEGRAB NA, WILLIAMS AC, BARRY BW: Oestradiol permeation through human skin silastic membrane: effects of propylene glycol and supersaturation. *J. Control. Rel.* (1995) **36**:277-294.
18. MA X, TAW J, CHIANG C: Control of drug crystallization in transdermal matrix systems. *Int. J. Pharm.* (1996) **142**:115-119.
19. SCHWARB FP, IMANIDIS NG, SMITH EW, HAIGH JM, SURBER C: Effect of concentration and degree of saturation of topical flucocinonide formulations on *in vitro* membrane transport and *in vivo* availability on human skin. *Pharm. Res.* (1997) **16**:909-915.
20. RAGHAVAN SL, TRIVIDIC A, DAVIS AF, HADGRAFT J: Crystallization of hydrocortisone acetate: influence of polymers. *Int. J. Pharm.* (2001) **212**:213-221.
- **Evaluation of HPMC, methyl cellulose, PVP and PEG 400 on crystallisation of hydrocortisone acetate from supersaturated solutions, showing HPMC was most effective in delaying crystallisation.**
21. RAGHAVAN RL, KIEPFER B, DAVIS AF, KAZARIAN SG, HADGRAFT J: Membrane transport of hydrocortisone acetate from supersaturated solutions; the role of polymers. *Int. J. Pharm.* (2001) **221**:95-105.
22. PELLET MA, DAVIS AF, HADGRAFT J: Effect of supersaturation on membrane transport: 2. Piroxicam. *Int. J. Pharm.* (1994) **111**:1-6.
23. PELLET MA, CASTELLANO S, HADGRAFT J, DAVIS AF: The penetration of supersaturated solutions of piroxicam across silicone membranes and human skin *in vitro*. *J. Control. Rel.* (1997) **46**:205-214.
24. PELLET MA, ROBERTS MS, HADGRAFT J: Supersaturated solutions evaluated with an intro stratum corneum tape stripping technique. *Int. J. Pharm.* (1997) **151**:91-98.
25. RAGHAVAN RL, TRIVIDIC A, DAVIS AF, HADGRAFT J: Effects of cellulose polymers on supersaturation and *in vitro* membrane transport of hydrocortisone acetate. *Int. J. Pharm.* (2000) **193**:231-237.
26. IERVOLINO M, RAGHAVAN RL, HADGRAFT J: Membrane penetration enhancement of ibuprofen using supersaturation. *Int. J. Pharm.* (2000) **198**:229-238.
27. IERVOLINO M, CAPPELLO B, RAGHAVAN RL, HADGRAFT J: Penetration enhancement of ibuprofen from supersaturated solutions through human skin. *Int. J. Pharm.* (2001) **212**:131-141.
28. SIMONELLI AP, MEHTA SC, HIGUCHI WI: Inhibition of sulfathiazole crystal growth by polyvinylpyrrolidone. *J. Pharm. Sci.* (1970) **59**:633-638.
29. SEKIKAWA H, FUJIWARA J, NAGANUMA T, NAKANO M, ARITA T: Dissolution behaviors and gastrointestinal absorption of phenytoin in phenytoin-polyvinylpyrrolidone coprecipitate. *Chem. Pharm. Bull.* (1978) **26**:3033-3039.
30. SUZUKI H, SUNADA H: Comparison of nicotiamide, ethylurea and polyethylene glycol as carriers for nifedipine solid dispersion systems. *Chem. Pharm. Bull.* (1997) **45**(10):1688-1693.
31. SUZUKI H, SUNADA H: Some factors influencing the dissolution of solid dispersions with nicotiamide and hydroxypropylmethylcellulose as combined carriers. *Chem. Pharm. Bull.* (1998) **46**(6):1015-1020.
32. YAMADA T, SAITO N, IMAI T, OTAGIRI M: Effect of grinding with hydroxypropyl cellulose on dissolution and particle size of a poorly water-soluble drug. *Chem. Pharm. Bull.* (1999) **47**(9):1311-1313.
33. KOHRI N, YAMAYOSHI Y, XIN H *et al.*: Improving the oral bio-availability of albendazole in rabbits by the solid dispersion technique. *J. Pharm. Pharmacol.* (1999) **51**:159-164.
- **HPMC and hydroxypropyl methylcellulose acetate phthalate inhibited the crystallisation of albendazole from supersaturated solutions.**
34. HASEGAWA A, NAKAGAWA H, SUGIMOTO I: Application of solid dispersions of nifedipine with enteric coating agent to prepare a sustained-release dosage form. *Chem. Pharm. Bull.* (1985) **33**(4):1615-1619.
35. O'DRISCOLL KM, CORRIGAN OI: Chlorothiazide-polyvinylpyrrolidone (PVP) interactions: influence on membrane permeation (everted rat intestine) and dissolution. *Drug Devel. Ind. Pharm.* (1982) **8**(4):547-564.
36. USUI F, MAEDA K, KUSAI A, NISHIMURA K, YAMAMOTO K: Inhibitory effects of water soluble polymers on precipitation of RS-8359. *Int. J. Pharm.* (1997) **154**:59-66.
37. HASEGAWA A, TAGUCHI M, SUZUKI R, MIYATA T, NAKAGAWA H, SUGIMOTO I: Supersaturation mechanism of drugs from solid dispersions with enteric coating agents. *Chem. Pharm. Bull.* (1988) **36**(12):4941-4950.
- **The rate of crystallisation of nifedipine, griseofulvin and spironolactone was remarkably inhibited by the presence of carboxymethylethyl cellulose. The inhibition was not due to solubilisation of the drugs, and it was suggested that crystal growth was inhibited due to adsorption of**

- carboxymethylethyl cellulose at the crystal surface.
38. GAO P, GUYTON ME, HUANG T, BAUER JM, STEFANSKI KJ, LU Q: Enhanced oral bioavailability of a poorly water soluble drug PNU-91325 by supersaturable formulations. *Drug Devel. Ind. Pharm.* (2004) **30**(2):221-229.
  39. GAO P, RUSH RD, PFUND WP *et al.*: Development of a supersaturable SEDDS (S-SEDDS) formulation of paclitaxel with improved oral bioavailability. *J. Pharm. Sci.* (2003) **92**(12):2395-2407.
  - **Enhanced free-drug level with the S-SEDDS formulation improves oral bioavailability.**
  40. LI LY, RODRIGUEZ-HORNEDO N, HEIMBACH, T, FLEISHER D: *In-vitro* crystallization of indinavir in the presence of ritonavir and as a function of pH. *J. Pharm. Pharmacol.* (2003) **55**(5):707-711.
  41. TANG L, KHAN SU, MUHAMMAD NA: Evaluation and selection of Bio-relevant dissolution media for a poorly water soluble new chemical entity. *Pharm. Dev. Tech.* (2002) **6**(4):531-540.
  - **Biorelevant dissolution media were developed and used in formulation development.**
  42. LEE J, LEE SC, ACHARYA G, CHANG C, PARK K: Hydrotropic solubilization of paclitaxel: analysis of chemical structures for hydrotropic property. *Pharm. Res.* (2003) **20**(7):1022-1030.
  43. STRAUDINGER RM: Chapter 9 Biopharmaceutics of paclitaxel (Taxol<sup>®</sup>): formulation, activity, and pharmacokinetics. In: *Taxol Science and Applications*. M Suffness (Ed.), CRC Press, Inc., New York, NY, USA (1995).
  44. In: *Handbook on Injectable Drugs (8th Edition)*. LA Trissel (Ed.) American Society of Health-System Pharmacists, Bethesda, MD, USA (1994):808.
  45. NUIJEN B, BOUMA M, SCHELLENS JH, BEIJNEN JH: Progress in the development of alternative pharmaceutical formulations of taxanes. *Inv. New Drugs.* (2001) **19**:143-153.
  46. ZUYLEN LV, VERWEIJ J, SPARREBOOM A: Role of formulation vehicles in taxane pharmacology. *Inv. New Drugs.* (2001) **19**:125-141.
  47. LASSUS M, SCOTT D, LEYLAND JB: Allergic reactions associated with Cremophor containing antineoplastics. *Proc. Am. Soc. Clin. Oncol.* (1985) **4**:268.
  48. MALINGRE MM, BEIJNEN JH, SCHELLENS JH: Oral delivery of taxanes. *Inv. New Drugs.* (2001) **19**:155-162.
  49. BARDELEIJER HA, TELLINGEN OV, SCHELLENS JH, BEIJNEN JH: The oral route of the administration of cytotoxic drugs: strategies to increase the efficiency and consistency of drug delivery. *Inv. New Drugs.* (2000) **18**:231-241.
  50. TERWOGT JM, MALINGRE MM, BEIJNEN JH *et al.*: Coadministration of oral cyclosporin A enables oral therapy with paclitaxel. *Clin. Cancer Res.* (1999) **5**:3379-3384.
  51. MALINGRE MM, BEIJNEN JH, ROSING H *et al.*: A Phase I and pharmacokinetic study of bi-daily dosing of oral paclitaxel in combination with cyclosporin A. *Cancer Chemother. Pharmacol.* (2001) **47**:347-354.
  52. WOO JS, LEE CH, SHIM CK, HWANG SJ: Enhanced oral bioavailability of paclitaxel by coadministration of the P-glycoprotein inhibitor KR30031. *Pharm. Res.* (2003) **20**:24-30.
  53. MONTASERI H, JAMALI F, MICETICH RG, DANESHTALAB M: Improving oral bioavailability of Taxol. *Pharm. Res.* (1995) **12**:S-429.
  54. GAO P, MOROZOWICH W: Design and development of supersaturable SEDDS (S-SEDDS) formulations for enhancing the gastrointestinal absorption of poorly soluble drugs. In: *Lipid Based Drug Delivery of Poorly Water Soluble Drugs*. D Hauss (Ed.), Marcel Dekker, New York, NY, USA (In press).
  55. DRESSMAN JB, REPPAS C: *In vitro-in vivo* correlations for lipophilic, poorly water-soluble drugs. *Eur. J. Pharm. Sci.* (2000) **11**(Suppl. 2):S73-S80.
  56. VOGELPOEL H, WELINK J, AMIDON GL *et al.*: Commentary: Biowaiver monographs for immediate release solid oral dosage forms based on biopharmaceutics classification system (BCS) literature data: verapamil hydrochloride, propranolol hydrochloride, and atenolol. *J. Pharm. Sci.* (2004) **93**(8):1945-1956.
  57. RAUB TJ, BARSUHN CL, WILLIAMS LR, DECKER DE, SAWADA GA, HO NF: Use of a biophysical-kinetic model to understand the roles of protein binding and membrane partitioning on passive diffusion of highly lipophilic molecules across cellular barriers. *J. Drug Target.* (1993) **1**(4):269-286.
  - **A theoretical analysis showing that the permeability of highly lipophilic molecules could decrease due to extensive association with the membrane.**
  58. KOSSENA GA, CHARMAN WN, BOYD BJ, PORTER CJ: Influence of the intermediate digestion phases of common formulation lipids on the absorption of a poorly water-soluble drug. *J. Pharm. Sci.* (2005) **94**(3):481-492.
  59. HE X, KADOMURA S, TAKEKUMA Y, SUGAWARA M, MIYAZAKI K: A new system for the prediction of drug absorption using a pH-controlled Caco-2 model: evaluation of pH-dependent soluble drug absorption and pH-related changes in absorption. *J. Pharm. Sci.* (2004) **93**(1):71-77.
  - **An *in vitro* dissolution system is described for the prediction of pH-dependent soluble drug absorption using the physiological conditions in the GI tract.**
  60. DELANEY B, STEVENS LA, SCHMELZER W *et al.*: Oral absorption of phytosterols and emulsified phytosterols by Sprague-Dawley rats. *J. Nutri. Biochem.* (2004) **15**(5):289-295.
  61. PORTER CJ, CHARMAN WN: *In vitro* assessment of oral lipid based formulations. *Adv. Drug Del. Rev.* (2001) **50**(Suppl. 1):S127-147.
  62. OSTLUND RE Jr, MCGILL JB, ZENG C *et al.*: Gastrointestinal absorption and plasma kinetics of soy D5-phytosterols and phytostanols in humans. *Am. J. Physiol.* (2002), **282**(4):E911-E916.
  63. BÖRGSTROM B, DAHLQUIST A, LUNDH G, SJÖVALL J: Studies of intestinal digestion and absorption in the human. *J. Clin. Invest.* (1957) **36**:1521-1529.
  - **First detailed study showing the role of the bile acid mixed micelle in the absorption of lipophilic substances.**
  64. BORGSTRÖM B, PATTON JS: Luminal events in gastrointestinal lipid digestion. In: *Handbook of Physiology Section 6. The Gastrointestinal System IV*. SG Schultz (Ed.), American Physiological Society, Bethesda, MD, USA (1991):475-504.
  - **Excellent review of the absorption of lipids and the role of the bile acid mixed micelle.**

65. STAGGERS JE, HERNELL O, STAFFORD RJ, CAREY MC: Physical-chemical behavior of dietary and biliary lipids during intestinal digestion and absorption. 1. Phase behavior and aggregation states of model lipid systems patterned after aqueous duodenal contents of healthy adult human beings. *Biochemistry* (1990) **29**:2028-2040.
- **Detailed study of intestinal lipid absorption.**
66. HERNELL O, STAGGERS JE, CAREY MC: Physical-chemical behavior of dietary and biliary lipids during intestinal digestion and absorption. 2. Phase analysis and aggregation states of luminal lipids during duodenal fat digestion in healthy adult human beings. *Biochem.* (1990) **29**:2041-2056.
- **Detailed study of intestinal lipid absorption.**
67. CAREY MC, SMALL DM: The characteristics of mixed micellar solutions with particular reference to bile. *Am. J. Med.* (1970) **49**:590-598.
68. HOFMANN AF, BORGSTRÖM B: The intraluminal phase of fat digestion in man: the lipid content of the micellar and oil phases of intestinal content obtained during fat digestion and absorption. *J. Clin. Invest.* (1964) **43**:247-257.
69. MATTSON FH, VOLPENHEIN RA: The digestion and absorption of triglycerides. *J. Biol. Chem.* (1964) **239**:2772-2777.
- **In the intestinal lumen, triglycerides are hydrolysed to monoglycerides and free glycerol with cleavage of all of the fatty acids esterified at the  $\alpha$ - and  $\alpha'$ -positions, and 22% of those esterified at the  $\beta$ -position of the dietary triglycerides. Of the glycerol, ~ 75% of dietary triglyceride is absorbed as monoglyceride, and 75% of the fatty acids are absorbed as free acids. The free glycerol is absorbed independently of the lipids, and little of it is used in glyceride synthesis. In the intestinal wall, the monoglycerides are re-esterified to triglycerides.**
70. SIMMONDS WJ, HOFMANN AF, THEODOR E: Absorption of cholesterol from a micellar solution: intestinal perfusion studies in man. *J. Clin. Invest.* (1967) **45**:874-890.
71. WILSON FA, DIETSCHY JM: Characterization of bile acid absorption across the unstirred water layer and brush border of the rat jejunum. *J. Clin. Invest.* (1972) **51**(12):3015-3025.
72. MATTSON FH, NOLEN GA: Absorbability by rats of compounds containing one to eight ester groups. *J. Nutr.* (1972) **102**:1171-1176.
73. GROVES MJ, MUSTAFA RM: Measurement of the 'spontaneity' of self-emulsifiable oils. *J. Phar. Pharmacol.* (1974) **26**:671-681.
74. MANSBACH CM, COHEN RS, LEFF PB: Isolation and properties of the mixed lipid micelles present in intestinal content during fat digestion in man. *J. Clin. Invest.* (1975) **56**:781-791.
- **Duodenal fluid was collected after a 36-g fat breakfast and the aqueous phase containing the mixed micelles (1.5 nm) showed a stoichiometry of 1.4 mol fatty acid, 0.15 mol lysolecithin and 0.06 mol cholesterol for each mole of bile acid.**
75. WESTERGAARD H, DIETSCHY JM: The mechanism whereby bile acid micelles increase the rate of fatty acid and cholesterol uptake into the intestinal mucosal cell. *J. Clin. Invest.* (1976) **58**:97-108.
76. HO NFH, PARK JY, MOROZOWICH W, HIGUCHI WI: Physical model approach to the design of drugs with improved intestinal absorption. In: *Design of Biopharmaceutical Properties Through Prodrugs and Analogs*. EB Roche (Ed.) APhA Academy of Pharmaceutical Sciences, Washington, DC, USA (1978):136-227.
77. MONTET C, REYNIER MO, MONTET AM, GEROLAMI A: Distinct effects of three bile salts on cholesterol solubilization by oleate-monoolein-bile salt micelles. *Biochim. Biophys. Acta.* (1979) **575**(2):289-294.
78. SHIAU YF, LEVINE GM: pH dependence of micellar diffusion and dissociation. *Am. J. Physiol.* (1980) **239**:G177-G182.
79. SAWADA GA, BARSUHN CL, LUTZKE BS *et al.*: Increased lipophilicity and subsequent cell partitioning decrease passive transcellular diffusion of novel, highly lipophilic antioxidants. *J. Pharmacol. Exp. Therap.* (1999) **288**(3):1317-1326.
- **Highly lipophilic compounds may show reduced permeation coefficient values.**
80. THOMSON AB, O'BRIEN BD: Uptake of cholesterol into rabbit jejunum using three *in vitro* techniques: importance of bile acid micelles and unstirred layer resistance. *Am. J. Physiol.* (1981) **241**:G270-G274.
81. IRANLOYE TA, PILPEL N, GROVES MJ: Some factors affecting the droplet size and charge of dilute oil-in-water emulsions prepared by self-emulsification. *J. Disp. Sci. & Tech.* (1983) **4**(2):109-121.
82. SHIAU YF: Lipid digestion and absorption. In: *Physiology of the Gastrointestinal Tract (2nd Ed.)*. LR Johnson (Ed.), Raven Press, New York, NY, USA (1987):1527-1556.
83. REYNIER MO, CROTTE C, MONTET JC, SAUVE P, GEROLAMI A: Intestinal cholesterol and oleic acid uptake from solutions supersaturated with lipids. *Lipids.* (1987) **22**(1):28-32.
- **Lipid absorption and intestinal uptake of fatty acids and cholesterol occurs via both the micellar saturated and nonmicellar supersaturated phases.**
84. POELMA FG, TUKKER JJ, CROMMELIN DJ: Intestinal absorption of drugs. I. The influence of taurocholate on the absorption of dantrolene in the small intestine of the rat. *J. Pharm. Sci.* (1989) **78**:285-289.
- **The intestinal permeation coefficient value is directly related to the free-drug concentration and not to the total concentration of the micellar phase and the free drug.**
85. THURNHOFER H, HAUSER H: Uptake of cholesterol by small intestinal brush border membrane is protein-mediated. *Biochem.* (1990) **29**:2142-2148.
- **Cholesterol undergoes collisional transfer from BA micelles with the intestinal brush border. The absorption of cholesterol by the small intestinal brush border membrane is either from mixed micelles or from small unilamellar vesicles. The kinetic data are consistent with a mechanism involving collisional-induced transfer of cholesterol.**
86. POELMA FG, BREAS R, TUKKER JJ, CROMMELIN DJ: Intestinal absorption of drugs. The influence of mixed micelles on the disappearance kinetics of drugs from the small intestine of the rat. *J. Pharm. Pharmacol.* (1991) **43**(5):317-324.
87. SMALL DM: The effects of glyceride structure on absorption and metabolism. *Ann. Rev. Nutr.* (1991) **11**:413-434.
- **Excellent review of triglyceride digestion, absorption, enterocyte transport and tissue accumulation of hydrolytic products.**
88. THURNHOFER H, SCHNABEL J, BETZ M, LIPKA G, PIDGEON C, HAUSER H: Cholesterol-transfer protein located in the intestinal brush border membrane. Partial purification and characterization. *Biochim. Biophys. Acta.* (1991) **1064**:275-286.

- **Collisional transfer of cholesterol occurs with the enterocyte brush border.**
- 89. LIPKA G, IMFELD D, SCHULTHESS G, THURNHOFER H, HAUSER H: Protein mediated cholesterol absorption by small intestinal brush border membranes. In: *Structural and Dynamic Properties of Lipids and Membranes* PG Quinn, RJ Cherry (Eds), Portland Press, London, UK (1992):7-18.
- 90. THOMSON AB, SCHOELLER C, KEELAN M, SMITH L, CLANDININ MT: Lipid absorption: passing through the unstirred layers, brush-border membrane, and beyond. *Can. J. Physiol. Pharm.* (1993) 71:531-555.
- **Comprehensive review of lipid absorption.**
- 91. TSO P: Intestinal lipid absorption. In: *Physiology of the Gastrointestinal Tract (3rd Edition)*, LR Johnson (Ed.), Raven Press, New York, NY, USA (1994):1867-1907.
- **Excellent review.**
- 92. SCHOELLER SC, KEELAN M, MULVEY G, STREMMEL W THOMSON AB: Oleic acid uptake into rat and rabbit jejunal brush border membranes. *Biochim. Biophys. Acta.* (1995) 1236:51-64.
- 93. NARAYANAN VS, STORCH J: Fatty acid transfer in taurodeoxycholate mixed micelles. *Biochem.* (1996) 35:7466-7473.
- 94. DAWSON A, RUDEL LL: Intestinal cholesterol absorption. *Curr. Opin. Lipidol.* (1999) 10:315-320.
- **Evidence supports a protein-mediated mechanism for cholesterol uptake into the intestinal mucosal cell.**
- 95. BOSNER SM, LANGE LG, STENSON WF, OSTLUND RE: Percent cholesterol absorption in normal women and men quantified with dual stable isotopic tracers and negative ion mass spectrometry. *J. Lipid. Res.* (1999) 40:302-308.
- 96. WANG DQ: New concepts of mechanism of intestinal cholesterol absorption. *Ann. Hepatol.* (2003) 2(3):113-121.
- 97. KRAMER W, GIRBIG F, CORSIERO D *et al.*: Intestinal cholesterol absorption: identification of different binding proteins for cholesterol and cholesterol absorption inhibitors in the enterocyte brush border membrane. *Biochim. Biophys. Acta.* (2003) 1633(1):13-26.
- 98. DELANEY B, STEVENS LA, SCHMELZER W *et al.*: Oral absorption of phytosterols and emulsified phytosterols by Sprague-Dawley rats. *J. Nutr. Biochem.* (2004) 15(5):289-295.
- **Plant sterols in the diet reduces intestinal uptake of cholesterol by reducing the intestinal micellar solubility and, hence, its transport towards the intestinal brush border membrane decreases.**
- 99. WIEDMANN TS, LIANG W, KAMEL L: Solubilization of drugs by physiological mixtures of bile salts. *Pharm. Res.* (2002) 19(8):1203-1208.
- 100. CHIJIWA K, LINSCHER WG: Effect of intraluminal pH on cholesterol and oleic acid absorption from micellar solutions in the rat. *Am. J. Physiol.* (1984) 246(5):G492-G499.
- 101. CHIJIWA K, LINSCHER WG: Mechanism of pH effect on oleic acid and cholesterol absorption in the rat. *Am. J. Physiol.* (1987) 252(4):G506-G510.
- **Low intestinal pH can reduce uptake of oleic acid.**
- 102. MISHKIN S, YALOVSKY M, KESSLER JI: Stages of uptake and incorporation of micellar palmitic acid by hamster proximal intestinal mucosa. *J. Lipid Res.* (1972) 13(2):155-68.
- 103. HORIUCHI K, NAITO I, NAKANO K *et al.*: Three-dimensional ultrastructure of the brush border glycocalyx in the mouse small intestine: a high resolution scanning electron microscopic study. *Arch. Histol. Cytol.* (2005) 68(1):51-6.
- **The glycocalyx on the luminal surface of the intestinal villi covered the top of the microvilli of the epithelial cells and the glycocalyx is observed as filamentous structures, 7 – 15 nm in diameter.**
- 104. BAKATSELOU V, OPPENHEIM RC, DRESSMAN JB: Solubilization and wetting effects of bile salts on the dissolution of steroids. *Pharm. Res.* (1991) 8(12):1461-1469.
- **Drug are solubilised by bile salts in a predictive fashion.**
- 105. TENHOOR CN, BAKATSELOU V, DRESSMAN JB: Solubility of mefenamic acid under simulated fed- and fasted-state conditions. *Pharm. Res.* (1991) 8(9):1203-1205.
- **Increased solubility of mefenamic acid in simulated fed-state media.**
- 106. MITHANI SD, BAKATSELOU V, TENHOOR CN, DRESSMAN JB: Estimation of the increase in solubility of drugs as a function of bile salt concentration. *Pharm. Res.* (1996) 13(1):163-167.
- **Poorly water soluble drugs can have high solubility in bile acid micelles.**
- 107. STORCH J, KLEINFELD AM: Transfer of long chain fluorescent free fatty acids between unilamellar vesicles. *Biochem.* (1986) 25(7):1717-1726.
- **Collisional transfer with different size populations of vesicles. Shorter chain fatty acids can equilibrate via the aqueous phase.**
- 108. CHO MJ, CHEN FJ, HUCZEK DL: Effects of inclusion complexation on the transepithelial transport of a lipophilic substance *in vitro*. *Pharm. Res.* (1995) 12(4):560-564.
- **The transepithelial transport of poorly soluble moderately lipophilic benzophenones (ClogP 2.6 – 4.1) is attributed to the passive diffusion of available free-drug molecules rather than the collision complex transfer at the enterocyte cell surface.**
- 109. HO SY, STORCH J: Common mechanisms of monoacylglycerol and fatty acid uptake by human intestinal Caco-2 cells. *Am. J. Physiol.* (2001) 281(4):C1106-C1117.
- 110. SCHULTHESS G, LIPKA G, COMPASSI S *et al.*: Absorption of monoacylglycerols by small intestinal brush border membrane. *Biochem.* (1994) 33(15):4500-4508.
- **The mechanism of the hexadecylglycerol absorption involves mainly monomer diffusion and probably collision-induced transfer.**
- 111. HSU KT, STORCH J: Fatty acid transfer from liver and intestinal fatty acid-binding proteins to membranes occurs by different mechanisms. *J. Biol. Chem.* (1996) 271(23):13317-13323.
- **These data strongly suggest that fatty acid transfer from intestinal fatty acid-binding proteins to membranes occurs by direct collisional interaction of the protein with the phospholipid bilayer.**
- 112. TROTTER PJ, HO SY, STORCH J: Fatty acid uptake by Caco-2 human intestinal cells. *J. Lipid Res.* (1996) 37(2):336-346.
- 113. HO SY, DELGADO L, STORCH J: Monoacylglycerol metabolism in human intestinal Caco-2 cells: evidence for metabolic compartmentation and hydrolysis. *J. Biol. Chem.* (2002) 277(3):1816-1823.
- 114. CONSTANTINIDES PP, SCALART JP: Formulation and physical characterization of water-in-oil microemulsions containing

- long- versus medium-chain glycerides. *Int. J. Pharm.* (1997) **158**:57-68.
115. HAUSS DJ, FOGAL SE, FICORILLI JV *et al.*: Lipid-based delivery systems for improving the bioavailability and lymphatic transport of a poorly water-soluble LTB4 inhibitor. *J. Pharm. Sci.* (1998) **87**:164-169.
116. HAQUE A, MORRIS ER: Thermogelation of methylcellulose. Part I. Molecular structures and processes. *Carbohydrate Polymers* (1993) **22**:161-173.
117. HAQUE A, RICHARDSON RK, MORRIS ER, GIDLEY MJ, CASWELL DC: Thermogelation of methylcellulose. Part II. Effect of hydroxypropyl substituents. *Carbohydrate Polymers* (1993) **22**:175-186.
118. ZILLER KH, RUPPRECHT H: Control of crystal growth in drug suspensions. *Drug Dev. Ind. Pharm.* (1988) **14**:2341-2370.

#### Website

201. <http://arbl.cvmb.colostate.edu/hbooks/contrib.html>  
AUSTGEN L, BOWEN RA, ROUGE M: *Pathophysiology of the digestive system*. Colorado State University, CO, USA (2004)

- See statement: 'As the ingesta is mixed, the bile salt mixed micelles *bump* into the brush border and the lipids, including monoglyceride and fatty acids, are absorbed.'

#### Affiliation

Ping Gao<sup>†</sup> PhD & Walter Morozowich PhD  
<sup>†</sup>Author for correspondence  
PGRD, Pfizer, Inc., 301 Henrietta Street,  
Kalamazoo, MI 49007, USA  
Tel: +1 269 833 6474; Fax: +1 269 833 2325;  
E-mail: ping.gao@pfizer.com; pgaous@yahoo.com